

Influence of Tumor Necrosis Factor- α on Insulin-like Growth Factor-I Binding Sites in Thecal Cells

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Story in Brief

The effect of bovine tumor necrosis factor- α (TNF α) on insulin-like growth factor-I (IGF-I) binding sites in follicular thecal cells of cattle was evaluated. Thecal cells were obtained from large (\approx 8 mm) ovarian follicles collected from cattle and cultured for 3 to 4 d. During the last 2 d of culture, cells were treated with recombinant bovine TNF α and specific binding of 125 I-IGF-I was determined. Two-day treatment with 10 and 30 ng/mL of TNF α decreased specific binding of 125 I-IGF-I to thecal cells. TNF α did not compete for 125 I-IGF-I binding to thecal cells whereas unlabeled IGF-I suppressed 125 I-IGF-I binding.

(Key Words: Tumor Necrosis Factor- α , Insulin-like Growth Factor-I, Thecal Cells, Cattle.)

Introduction

Tumor necrosis factor- α (TNF α) is a cytokine produced by activated macrophages, and in cattle systemic levels of TNF α increase acutely during infections (Kenison et al., 1990; Elsasser et al., 1995). It has been hypothesized that this increased level of TNF α may cause ovarian pathology such as ovarian cysts because TNF α directly inhibits granulosa and thecal cell steroidogenesis in cattle (Spicer and Alpizar, 1994; Spicer et al., 1995; Spicer, 1998). Alone, TNF α has little effect on ovarian steroidogenesis but in the presence of insulin-like growth factor-I (IGF-I), TNF α has dramatic inhibitory effects on steroidogenesis (Spicer, 1998). How TNF α accomplishes its inhibitory effect on steroidogenesis is unknown. Therefore, we set out to determine if the inhibitory effect of TNF α was due to either 1) a reduction in the number of IGF-I binding sites or 2) direct competition for IGF-I binding sites on bovine thecal cells.

Materials and Methods

Ovaries of beef and dairy cattle obtained at slaughter from a nearby abattoir were brought to the laboratory on ice and processed as previously described for obtaining thecal cells (Stewart et al., 1995). The medium was a 1:1 mixture of Dulbecco's modified Eagles medium and Ham's F12 containing gentamicin, glutamine and sodium bicarbonate. Approximately 2×10^5 viable cells were seeded in each plastic multiwell containing 1 ml of medium. Cultures were kept at 38.5° C in a 95% air – 5% CO₂ atmosphere, and medium was changed every 24 h. Cells were maintained in the presence of 10% fetal calf serum (FCS) for the first 2 d of culture, washed and treated for an additional 2 d with various doses (0, 3, 10 or 30 ng/ml) of recombinant bovine TNF α in serum-free medium for Experiment 1. For Experiment 2, cells were maintained in 10% FCS for a total of 3 d. At the end of the culture period, 125 I-IGF-I binding assays were conducted as previously described (Spicer and Stewart, 1996). In parallel cultures, numbers of cells were determined using a Coulter counter. Experimental data are presented as means \pm SE of measurement from replicated experiments. Each experiment was replicated at least three times. Binding of 125 I-IGF-I to thecal cells was expressed as cpm per 10^5 cells.

Results

In Experiment 1, 2-d treatment of thecal cells with 3 ng/mL of TNF α had no effect ($P > .10$) on specific binding of 125 I-IGF-I to thecal cells (Figure 1). However, 10 and 30 ng/ml of TNF α decreased ($P < .05$) specific binding of 125 I-IGF-I to thecal cells by 20% and 25%, respectively (Figure 1).

In Experiment 2, 10 and 100 ng/well of TNF α did not inhibit binding of ¹²⁵I-IGF-I to thecal cells, whereas both 10 and 100 ng/well of IGF-I inhibited binding of ¹²⁵I-IGF-I (Figure 2).

Discussion

Previous studies have shown that, in cattle, TNF α inhibits insulin- and IGF-I-induced estradiol production by granulosa cells and androstenedione production by thecal cells (Spicer and Alpizar, 1994; Spicer, 1998). Results of the present experiments indicate that TNF α may directly decrease the number of thecal cell IGF-I binding sites but that TNF α can not compete for IGF-I binding sites on thecal cells. Thus, the inhibitory effect of TNF α on IGF-I-induced thecal steroidogenesis may involve a decrease in the number of IGF-I receptors. However, the TNF α -induced decrease in IGF-I receptor numbers is much smaller (i.e., 20-25% decrease) than the TNF α -induced decrease in androstenedione production (i.e., 50% decrease) previously reported (Spicer, 1998) indicating that the inhibitory effect of TNF α on steroidogenesis may not be solely due to a TNF α effect on IGF-I receptor numbers. Additional studies will be required to clarify this possibility.

Previous studies have also shown that bovine thecal cells have high affinity, specific TNF α binding sites (Spicer, 1998). Thus, TNF α likely inhibits thecal steroidogenesis via signal transduction from its own receptor. The precise mechanism of action of TNF α on bovine thecal cell steroidogenesis awaits further elucidation. It has been suggested that TNF α may play an endocrine role and inhibit ovarian function during infection and disease (Spicer, 1998) and this may be why metritis increases the risk of abnormal estrus and cystic ovaries in lactating dairy cattle (Erb et al., 1985; Klerx and Smolders, 1997). However, this suggestion awaits verification.

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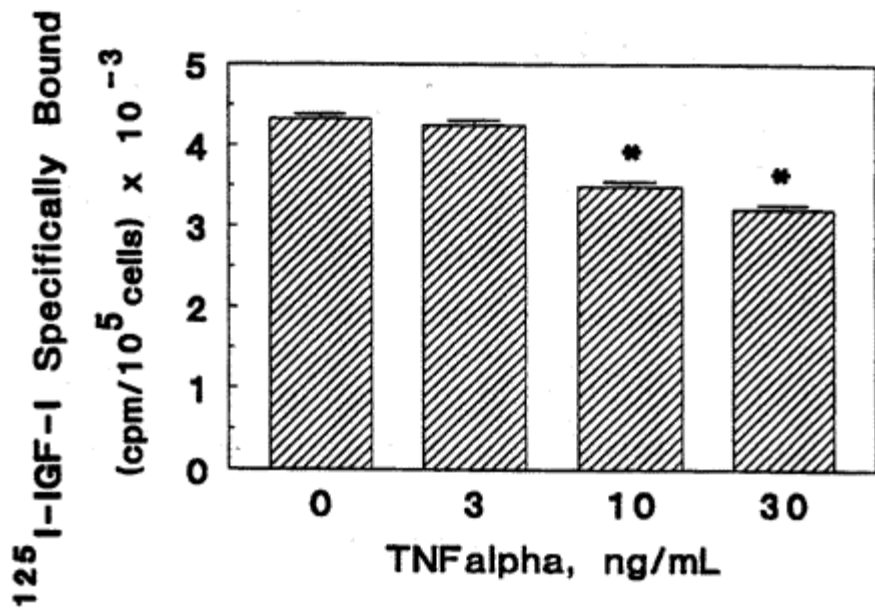


Figure 1. Effect of 2-d treatment of bovine TNF α on specific ¹²⁵I-IGF-I binding sites in bovine thecal cells (Experiment 1). After the 2-d treatments, cells were washed and then incubated with 100,000 cpm of ¹²⁵I-IGF-I. Non-specific binding was determined in the presence of 500 ng/well of recombinant human IGF-I. Values are means \pm SE of four separate experiments. *Mean differs ($P < .05$) from control (0 ng/ml).

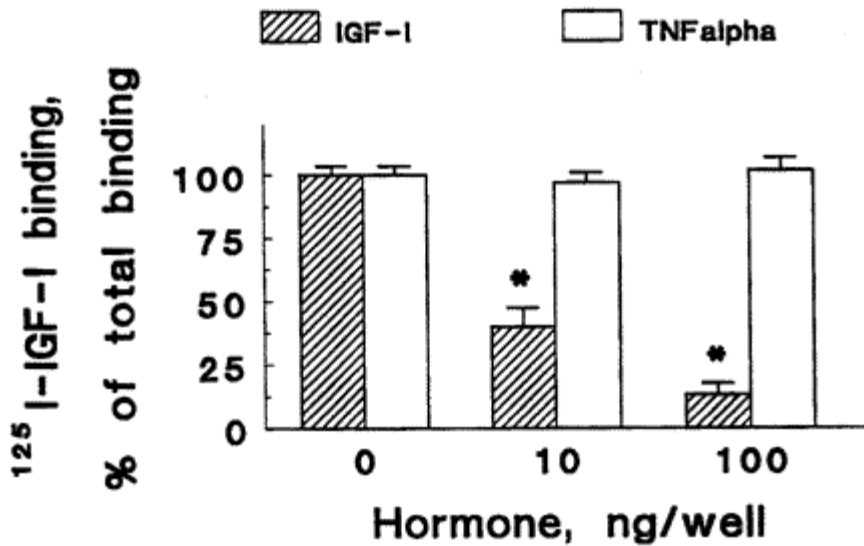


Figure 2. Comparison of TNF α and IGF-I on competing for IGF-I binding by thecal cells (Experiment 2). Thecal cells were cultured for 3 days in the presence of 10% FCS, and then cells were washed and incubated with 50,000 cpm of ¹²⁵I-IGF-I in the absence or presence of 0, 10 or 100 ng/well of TNF α or IGF-I. Values are means of three separate experiments and are expressed as a percentage of control total ¹²⁵I-IGF-I binding. *Mean

differs ($P < .05$) from control (0 ng/well).

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